

Suven Microdialysis Services

***In-Vivo Brain* Microdialysis Studies in Rodents for
Monitoring Changes in Neurotransmitters**

Acetylcholine

Histamine and Metabolite

GABA and Glutamate

Monoamines (NE, DA, 5HT & Metabolites)

**Simultaneous Monitoring of Neurotransmitters and Unbound
Test Compound Concentrations**

***In-Vivo Spinal* Microdialysis in Rodents for
Monitoring Changes in Inflammatory Biomarkers and
Neurotransmitters Responsible for Pain**

Mechanism of Action and PK/PD Studies

CSF Pharmacokinetics

Neurodegenerative models

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Suven Life Sciences Limited

Suven Life Sciences Limited is a Bio-pharmaceutical Company in existence since 1989 based at Hyderabad, India. Suven Drug Discovery & Development Support Services (DDDSS) is providing research services to global Pharma and Biotech companies.

Suven *In-Vivo* Microdialysis

List / Type of Studies being offered

Microdialysis team has expertise in conducting brain and spinal microdialysis studies to measure neuronal changes in freely moving animals.

Using brain microdialysis, we evaluate the modulation of neurotransmitters (and/or test compound) in most of the brain areas of freely moving animals.

Spinal microdialysis evaluates modulation of inflammatory biomarkers and neurotransmitters in spinal regions.

Neurotransmitters and test compounds are quantified by specific and sensitive analytical techniques.

Infrastructure

- Six Microdialysis Workstations
- Twelve Animals Can Undergo Dialysis Simultaneously
- Automated and Programmed Sample Collection under Refrigeration Over 24 hours
- Well Trained and Team of Scientists
- 2-3 Weeks Turnover Time for a Typical Study

Preclinical Species

- Rat (Wistar/Sprague Dawley)
- Guinea pig (Dunkin Hartley)
- Mouse

Neurotransmitters

- Acetylcholine
- Glutamate
- Gamma-amino butyric acid (GABA)
- Histamine
- Dopamine, Norepinephrine and Serotonin
- Monoamine Metabolites

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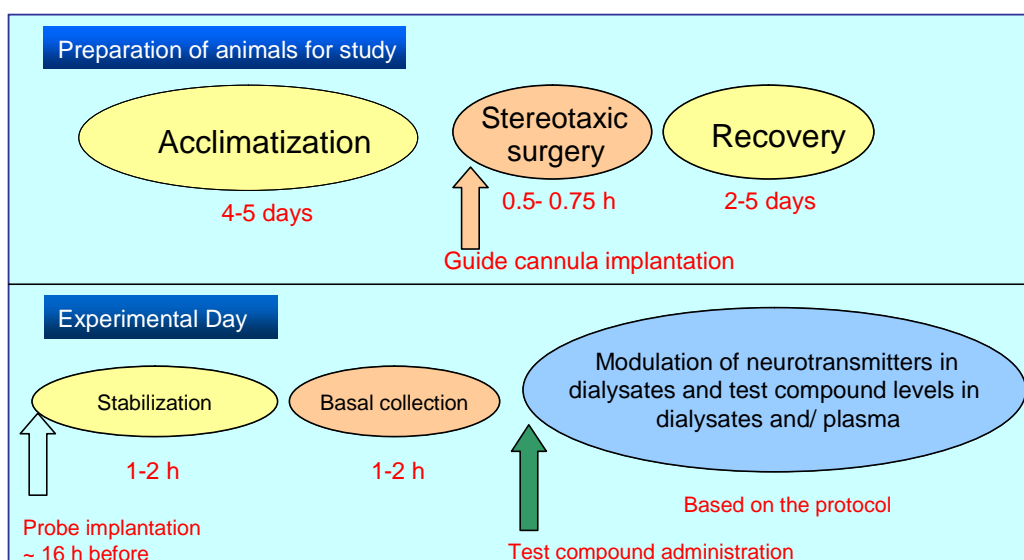
Study Designs

- Parallel Treatment Groups
- Cross-over Treatment Design
- Mechanism of Action Studies using various Antagonists/Blockers
- Pharmacokinetic, Pharmacodynamic (PK/PD) Studies *in vivo*, by simultaneous monitoring of neurotransmitters and drug concentrations

Routes of administration

- Systemic (*p.o.*, *i.p.*, *s.c.*, *i.v.* – bolus and infusion)
- Prolonged infusion using osmotic infusion pumps
- Local application (retrodialysis)
- Intracerebroventricular (ICV) injection

Typical experimental protocol



Flow rate: 1.0 – 2.0 $\mu\text{L}/\text{min}$

Sample collection intervals:

- Microdialysates: 15 - 30 min.
- Blood sampling (through jugular vein) from same animal: 6 – 8 points during absorption, distribution and elimination phase (or) same animal will be used after wash out period for exposure in plasma, brain and CSF (based on neurotransmitter profile)

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Fractions collectors:

- Samples are collected at 4 °C using programmed, refrigerated fraction collectors (up to 24 h post treatment).

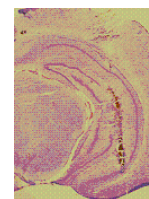


Histological Probe placement verification:

At the end of the experiment, probe placement is verified in each brain. Representative probe placement tracks in mPFC (a) and ventral hippocampus (b) of rat respectively.



(a)



(b)

Overview of Analytical Methods:

▪ Microdialysis samples:

- Acetylcholine : LC-MS/MS (API-4000)
- Amino acids : Glutamate and GABA – HPLC + Fluorescence
- Histamine : HPLC + Fluorescence
- Monoamines : LC-MS/MS (API-4000 Q-Trap)
- Monoamine metabolites : HPLC-ECD (BASi Epsilon)
- Test compounds : LC-MS/MS (API-4000 Q-Trap)
- Prostaglandin E2 : ELISA

▪ CSF samples:

- Monoamines (5-HT & 5-HIAA) : HPLC + ECD (BASi or ESA)
- Histamine and alpha methyl histamine : HPLC + Fluorescence
- Prostaglandin E2 : ELISA
- Interleukin-1 β : ELISA

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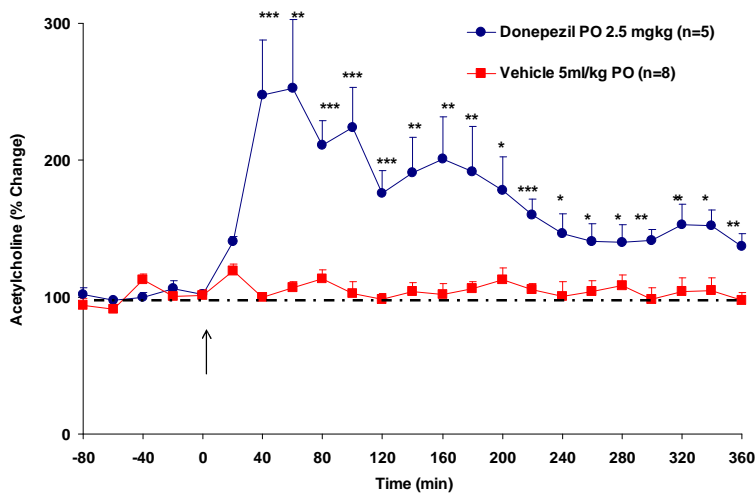
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- Plasma and Brain tissue samples:
 - Monoamines : HPLC + ECD (BASi or ESA)
 - Test compound : LC-MS/MS (API-4000 Q-Trap)
 -

Summary of Validation Experiments

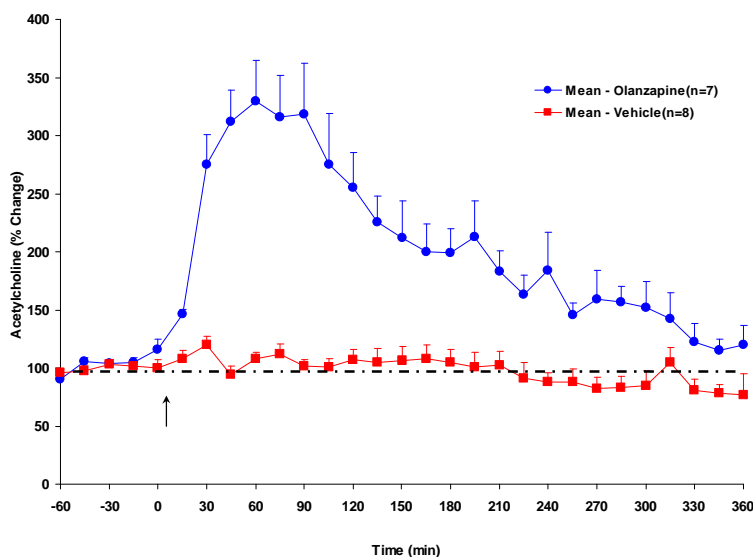
Acetylcholine

i. Modulation of acetylcholine in ventral hippocampus by donepezil in male Wistar rats



Perfusion fluid: aCSF
Flow rate: 1.5 μ L/min
Sampling duration: 20 min

ii. Modulation of acetylcholine in ventral hippocampus by Olanzapine in male Wistar rats



Perfusion fluid: aCSF
containing 0.1 μ mol neostigmine
Flow rate: 2.4 μ L/min
Sampling duration: 15 min

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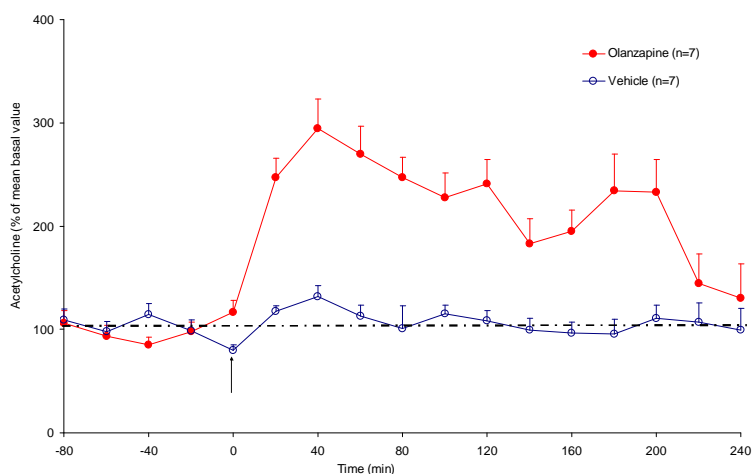
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iii. Modulation of acetylcholine by Olanzapine in prefrontal cortex of Dunkin-Hartley guinea pigs

pigs



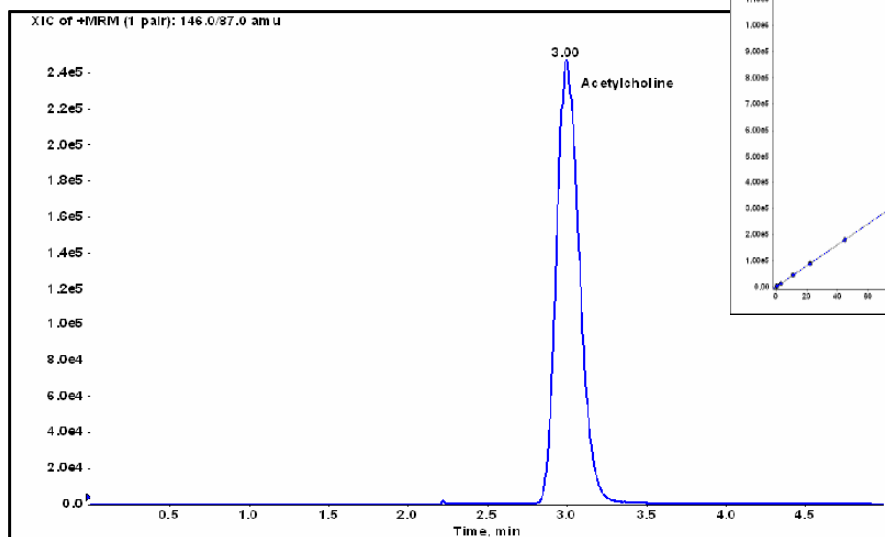
Perfusion fluid: aCSF containing 0.1 μ mol neostigmine
Flow rate: 1.5 μ L/min
Sampling duration: 20 min

Quantification of Acetylcholine

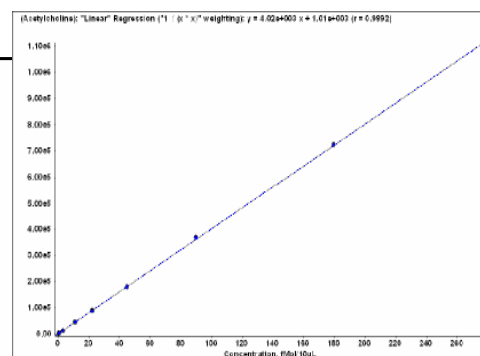
Acetylcholine concentrations in microdialysate samples are quantified by using HPLC coupled with tandem mass spectrometry (LC-MS/MS) without any sample pretreatment.

- Quantitation range: 0.05-103.50 nM

Typical chromatogram of acetylcholine



Calibration curve



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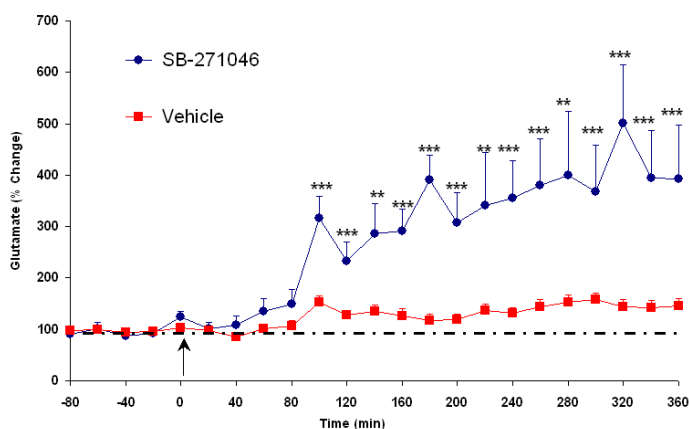
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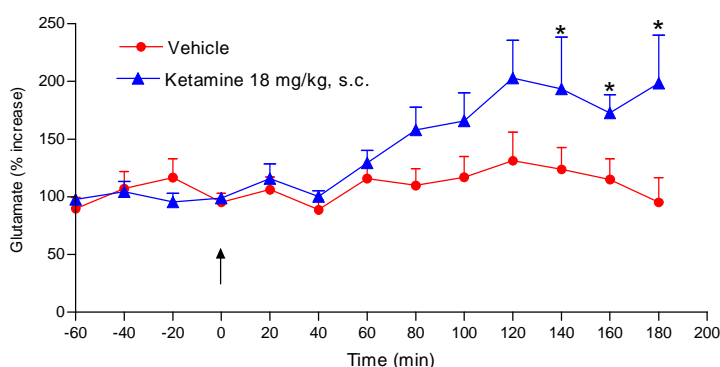
Glutamate

i. SB-271046: modulation of glutamate in frontal cortex of male Sprague-Dawley rats



Perfusion fluid: aCSF
Flow rate: 1.25 µL/min
Sampling duration: 15 min

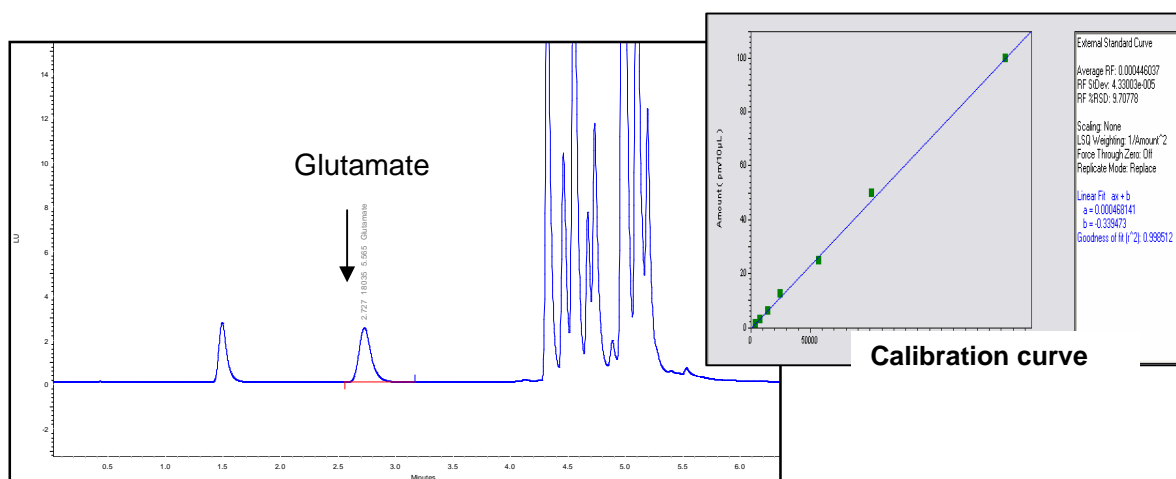
ii. Ketamine: modulation of glutamate in prefrontal cortex of male Sprague-Dawley rats



Perfusion fluid: aCSF
Flow rate: 3.0 µL/min
Sampling duration: 20 min

Quantification

Concentrations of glutamate in dialysates are determined by pre-column derivatization using HPLC-fluorescence method.



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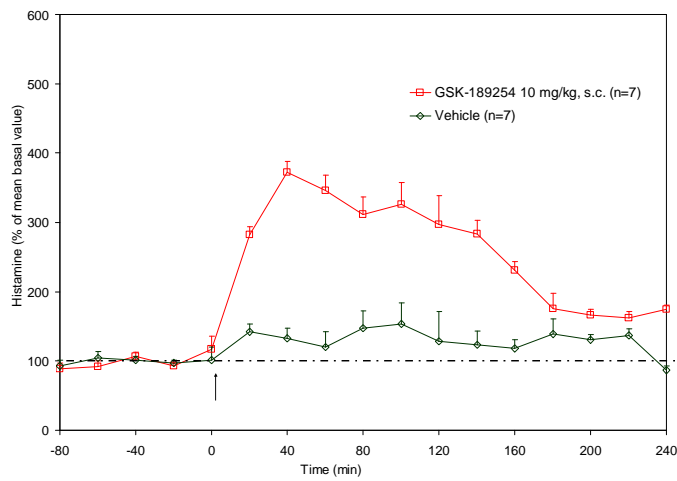
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Histamine

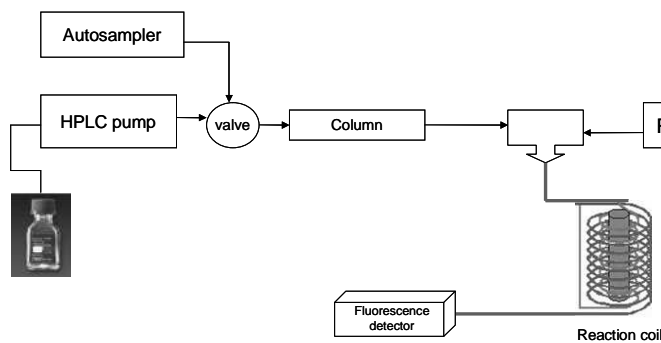
Modulation of histamine by GSK-189254 in prefrontal cortex of male Sprague-Dawley rats



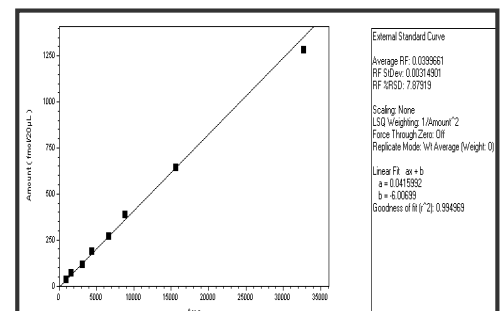
Perfusion fluid: aCSF
Flow rate: 1.5 μ L/min
Sampling duration: 20 min

Quantification

Concentrations of histamine in microdialysates are analyzed by HPLC and fluorometric detection after post-column derivatization with O-phthalaldehyde (OPA) reagent, which is delivered by a secondary flow system.



Calibration curve

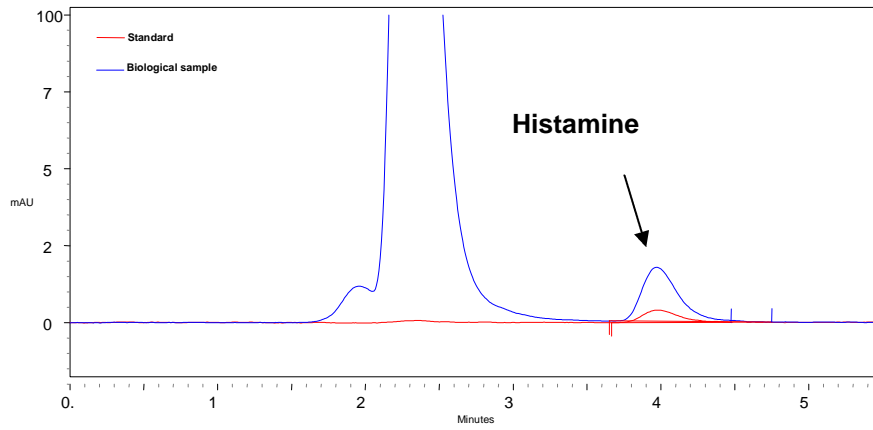


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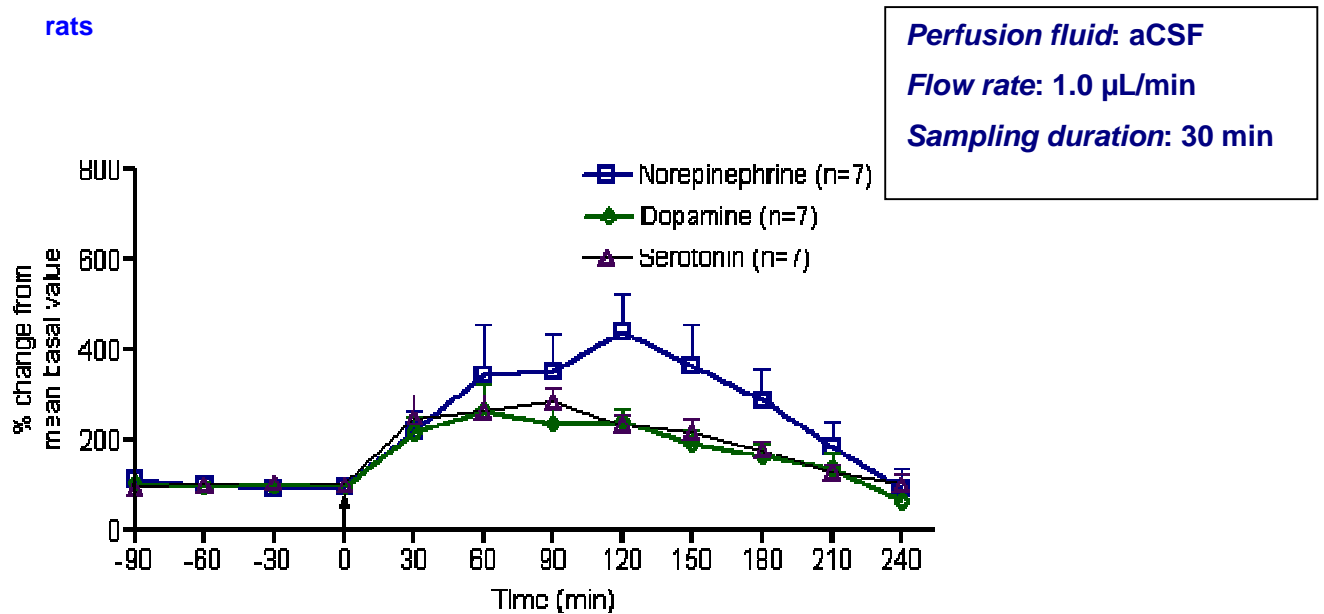
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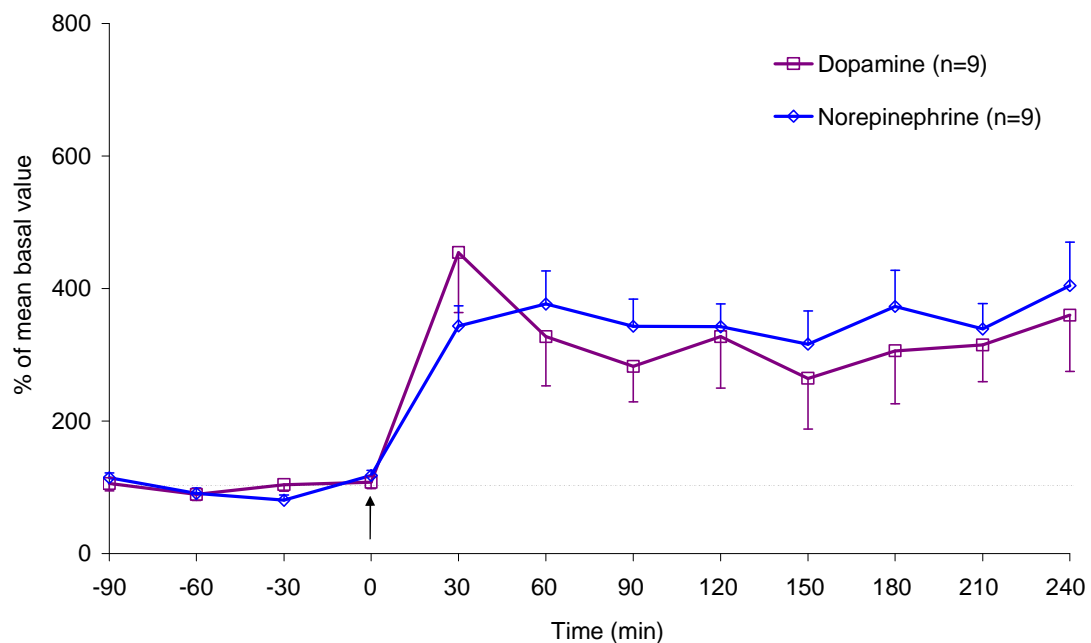
Dopamine, Norepinephrine and Serotonin

i. Modulation of monoamines by **venlafaxine** in prefrontal cortex of male **Sprague-Dawley**

rats



ii. Modulation of monoamines by **atomoxetine** in prefrontal cortex of male **Wistar** rats



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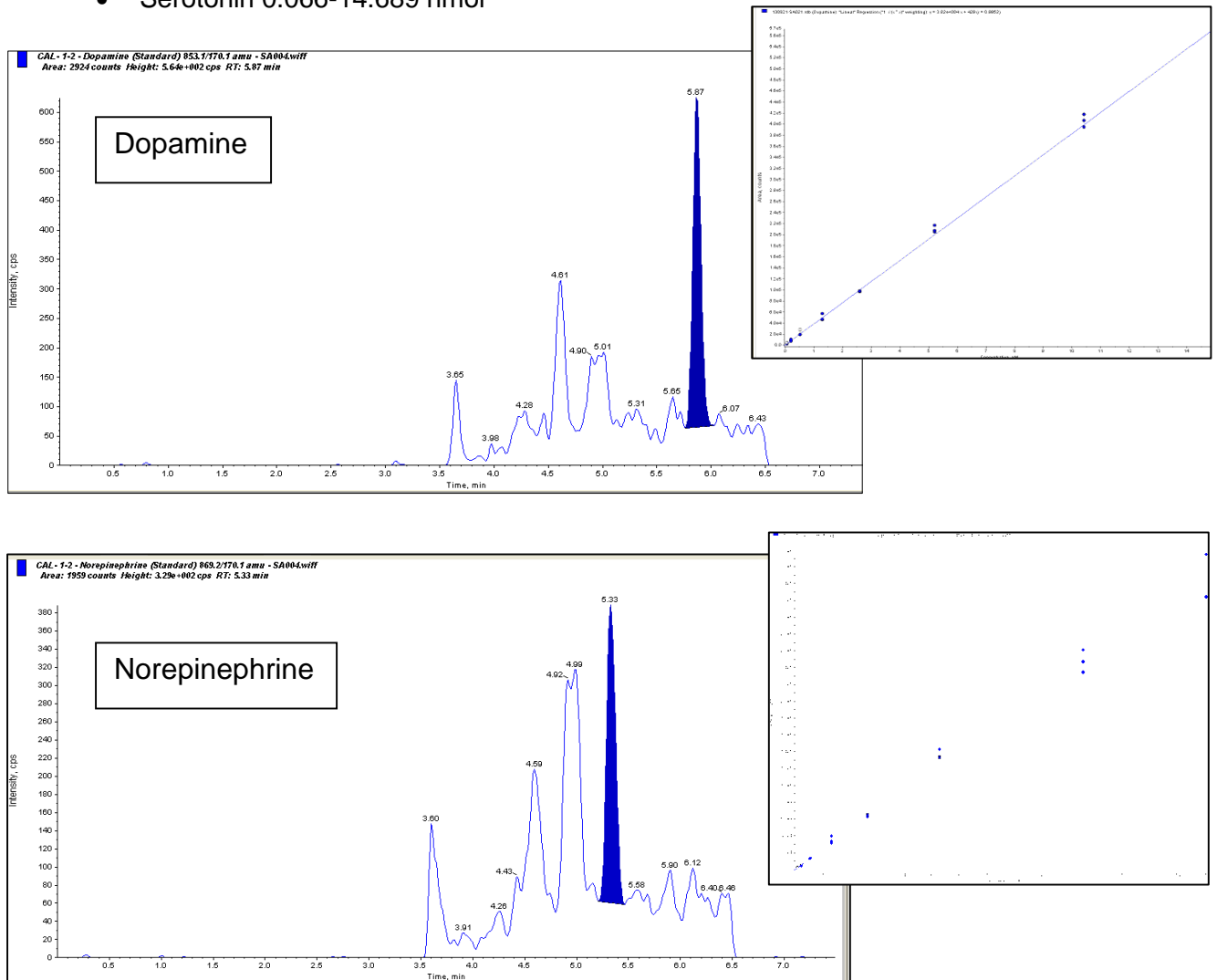
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Quantification

The catecholamine neurotransmitters dopamine, norepinephrine and serotonin were subjected for derivatization with dansyl chloride and following precursor-product ion pairs were monitored with m/z 853.1–170.1, m/z 869.2–170.1 and m/z 643.3-170.1 for the dansylated dopamine, norepinephrine and serotonin respectively. The analytes were quantified using triple quadrupole tandem mass spectrometer in positive ionisation mode using atmospheric pressure ionization source (Ref: Nirogi et al. 2013 Journal of Chromatography B. 913-914, p. 41-47). The test samples were quantified against a calibration curve prepared for each of the neurotransmitter using artificial cerebrospinal fluid in the calibration range of

- Dopamine 0.066-14.835 nmol
- Norepinephrine 0.066-14.728 nmol
- Serotonin 0.066-14.689 nmol

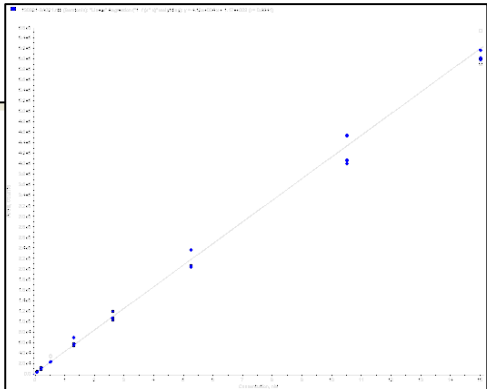
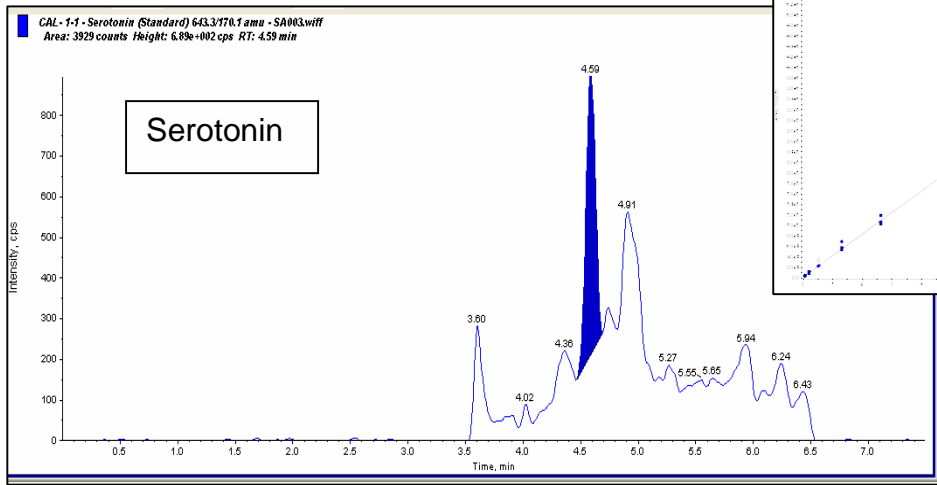


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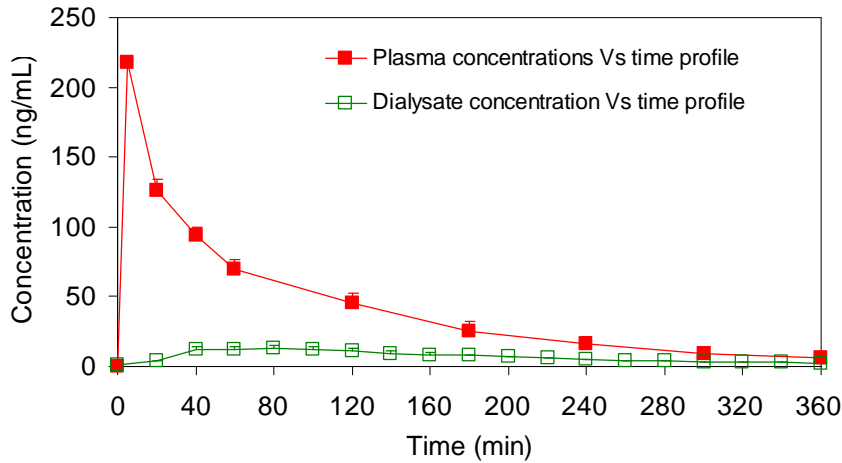
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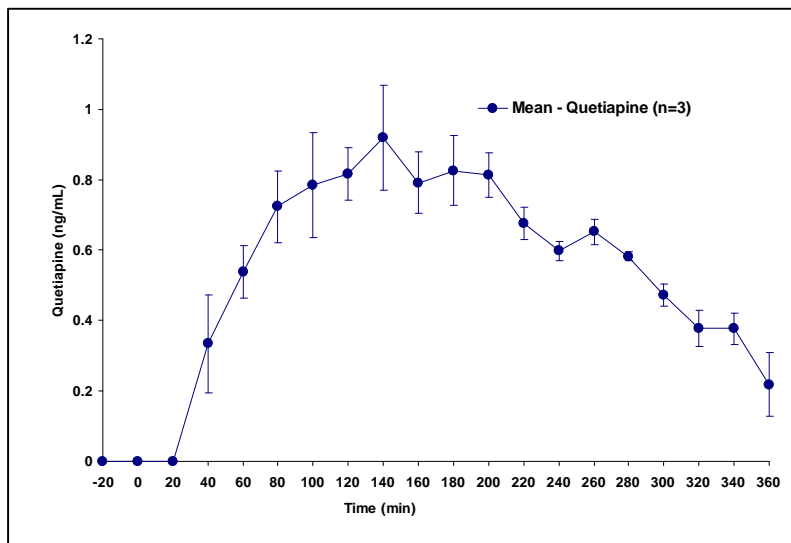
Determination of unbound brain concentrations of test compounds in rats.

i. Unbound concentration of escitalopram in prefrontal cortex



Perfusion fluid: aCSF
Flow rate: 1.0 $\mu\text{L}/\text{min}$
Sampling duration: 20 min
Test compound:
Escitalopram 2.5 mg/kg, i.v.

ii. Unbound brain concentration of quetiapine in striatum



Perfusion fluid: aCSF
Flow rate: 1.2 $\mu\text{L}/\text{min}$
Sampling duration: 20 min
Test compound: Quetiapine
30.0 mg/kg, s.c.

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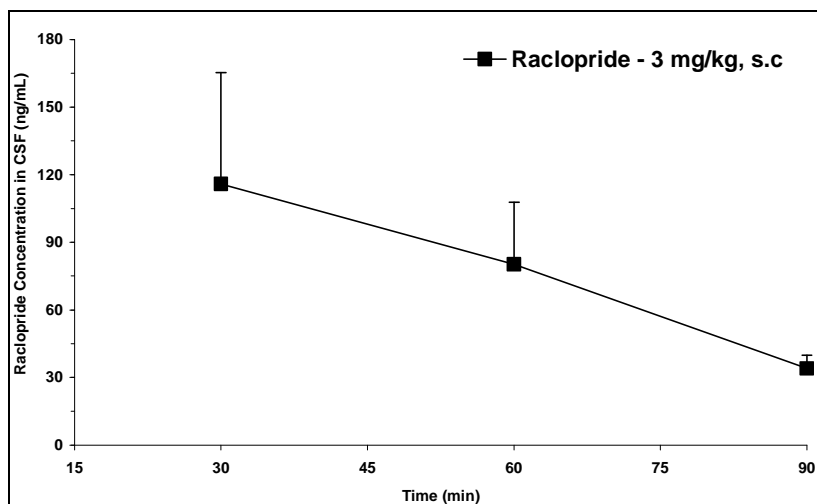
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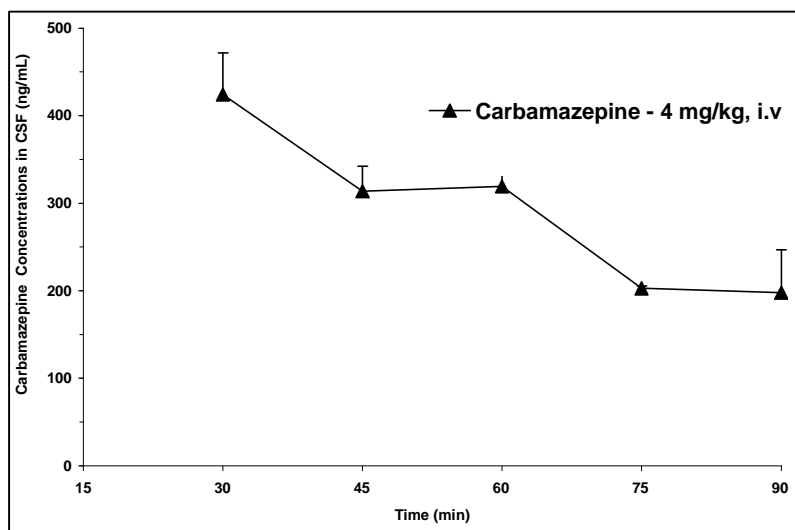
CSF and Brain Tissue Pharmacokinetic

i. CSF concentrations of Raclopride



Group size: n=5/ time point
Test compound: Raclopride
5.0 mg/kg, i.p.

ii. CSF concentrations of Carbamazepine



Group size: n=5/ time point
Test compound:
Carbamazepine 4.0 mg/kg,
i.v.

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